Association of *LPIN1* gene variations with markers of metabolic syndrome in population from Bosnia and Herzegovina

Tamer Bego¹, Tanja Dujić¹, Barbara Mlinar², Sabina Semiz^{1,4}, Maja Malenica¹, Besim Prnjavorac^{3,6}, Barbara Ostanek², Janja Marc², Anida Čaušević-Ramoševac⁵, Adlija Čaušević¹

¹Department of Biochemistry and Clinical Analysis, Faculty of Pharmacy, University of Sarajevo, Sarajevo, Bosnia and Herzegovina, ²Department of Clinical Biochemistry, Faculty of Pharmacy, University of Ljubljana, Ljubljana, Slovenia, ³General Hospital of Tešanj, Tešanj, ⁴Faculty of Engineering and Natural Sciences, International University of Sarajevo, Sarajevo, ⁵Bosnalijek, Pharmaceutical and Chemical Industry, Joint Stock Company, Sarajevo, ⁶Department of Pathophysiology, Faculty of Pharmacy; University Sarajevo, Sarajevo; Sarajevo; Bosnia and Herzegovina

ABSTRACT

Aim To investigate association of two *LPIN1* gene variations with main traits of metabolic syndrome (MS) (waist circumference, body mass index, blood pressure, triglycerides, HDL-cholesterol and fasting glucose levels) in population from Bosnia and Herzegovina.

Methods This study included 43 patients with metabolic syndrome and 43 healthy controls from General Hospital in Tešanj, Bosnia and Herzegovina. Subjects were genotyped for two *LPIN1* gene variations (rs11693809: C>T and rs2716610: C>T) by real time PCR method.

Results In control subjects *LPIN1* polymorphism, rs2716610: C>T, was significantly associated with a lower body mass index (BMI) (p=0.008) and waist circumference (p=0.008). The second analyzed rs11693809: C>T polymorphism was associated with lower blood HbA1c levels (p=0.048) in a group of MS patients.

Conclusion Results of our study suggest that rs2716610: C>T polymorphism of *LPIN1* gene could have a protective effect against development of metabolic syndrome, while rs11693809: C>T might affect a glucose control in patients with MS.

Keywords: metabolic syndrome, *LPIN1* gene, markers, gene variations.

Corresponding author:

Bosnia and Herzegovina

Fax: +387 33 586 188;

Phone: +387 33 586 188;

E.mail: tamer.bego@gmail.com

Department for Biochemistry and Clinical

Analysis, Faculty of Pharmacy, University

Zmaja od Bosne 8, 71000 Sarajevo,

Tamer Bego

of Sarajevo

Original submission: 28 November 2015; Revised submission: 03 February 2015; Accepted: 21 March 2015.

Med Glas (Zenica) 2015; 13(2):

INTRODUCTION

The metabolic syndrome (MS) is also known as syndrome X(1), the insulin resistance syndrome (2) and the deadly quartet (3). Metabolic syndrome is a combination of medical disorders that, when occurring together, increase the risk of developing cardiovascular disease and diabetes (4). The pathogenesis of the metabolic syndrome is thought to involve a complex interaction of multiple factors, including obesity and abnormal fat distribution, insulin resistance, hepatic, vascular, and immunologic factors, as well as lifestyle and genetic contributions (4). The available evidence indicates that between 20% and 30% of the world's adult population have MS (5). New harmonized definition and criteria for MS was accepted finally in 2009, which included measurement of waist circumference, blood pressure, triglycerides, HDL-cholesterol and serum glucose (6). Insulin resistance and abdominal obesity appear to be predominant underlying risk factors of MS. Beside this, other associated conditions for this syndrome can be genetic factors, physical inactivity, aging, and hormonal imbalance (7).

A newly discovered gene for lipin 1 (LPINI) resides in the 2p25 region, and codes for phosphatidic acid phosphatase, a key enzyme in triglyceride (TG) biosynthesis (8). There is evidence that chromosome region 2p25 is in linkage disequilibrium with several obesity related phenotypes, such as body mass index (BMI), waist circumference, skin-fold thickness, and percentage of body fat (9). The lipin protein family consists of three members, lipin-1, lipin-2, and lipin-3 (10). Lipin 1 is a newly discovered multifunctional protein that participates in the metabolism of lipids in different ways (11). Lipin-1 is abundantly expressed in adipose tissue and skeletal muscle, and lipin-1 protein localizes to either the cytosol or the nucleus, which may be related to its two known functions (12). Namely, in the cytosol, lipin-1 acts as a phosphatidate phosphatase (PAP) enzyme converting phosphatidate to diacylglycerol during triglyceride biosynthesis (13-14), while in the nucleus of adipocytes and hepatocytes lipin-1 acts as a transcriptional coactivator that interacts with the nuclear receptor peroxisome proliferator-activated receptor α (PPAR α) and PPAR γ (PPARG) coactivator 1α (PPARGC1 α) in a complex that modulates fatty acid oxidation gene expression (10, 15-16).

Null mutations in the murine Lpin1 gene result in a severe defect in adipose tissue development, which is related to insulin resistance and fatty liver dystrophy (fld mice) (10). Based upon the comprehensive biological knowledge of lipin-1 role in energy metabolism, human lipin-1 has been considered as an obvious biological candidate to explain some of the inter-individual variations in the common metabolic phenotypes. In addition, variants in LPIN1 have been associated with the fasting serum insulin levels, body mass index (BMI), waist circumference, and obesity development (17-19). Previous data from four meta-analysis studies found an association of LPIN1 variants with increased BMI (20), while another study found an association with hypertension (21). Other studies have shown correlation of expression of LPIN1 gene in adipose tissue with BMI and insulin resistance in humans (14, 16). Recently published German population study (n=1674), showed an interesting association of LPIN1 gene variants with metabolic phenotype (22). They identified three associated three-marker haplotypes, one common haplotype that increased the risk for metabolic syndrome, while other two were associated with lower blood pressure levels, lower BMI, waist circumference, and HbA₁C levels (22).

In this study, for the first time in population from Bosnia and Herzegovina, it was analyzed whether *LPIN1* gene polymorphisms (rs11693809: C>T and rs2716610: C>T), including haplotype analysis, were associated with the traits of metabolic syndrome. An association between biochemical parameters including, but not limited to, glucose, HbA1c, insulin levels, HDL and LDL cholesterol, triglycerides, serum proteins levels, and activity of liver enzymes and these two polymorphisms in *LPIN1* gene was analyzed in patients with metabolic syndrome and healthy controls.

PATIENTS AND METHODS

Study participants

The study included 43 patients with metabolic syndrome and 43 healthy controls from General Hospital in Tešanj, Zenica-Doboj Canton, Bosnia and Herzegovina. Investigation was done in accordance with ethical recommendations and practices of the General Hospital Tešanj, and with ethical principles outlined in the World Medical Association Declaration of Helsinki - Ethical Principles of Medical Research Involving Human Subjects (initiated in June 1964, last amendment in October 2000). Each subject in the study signed written informed consent. Metabolic syndrome was diagnosed according to new harmonized definition and criteria for MS from 2009. According to this new definition, MS is diagnosed when any three of the following five criteria are met: increased waist circumference (recommended waist circumference thresholds for Euripides) \geq 94 cm for men, and \geq 80 cm for women), triglycerides \geq 1.7 mmol/L, HDL-cholesterol < 1.0 mmol/L in males and < 1.3 mmol/L in females, blood pressure $\geq 130/85$ mmHg and fasting glucose \geq 5.6 mmol/L (6). Patients treated with insulin and patients with acute infection and / or inflammation and endocrine disorders were excluded from the study. All patients included in the study were using heterogeneous therapy (74% of all patients received antihypertensive therapy, 58% were treated with glucose-lowering drugs, and 47% were treated with lipid-lowering drugs). Healthy control group consisted of 43 non-obese, age-matched subjects, who had less than three features of MS. They were not taking any medication during the course of the study.

Biochemical and anthropometrical measurements

Waist circumference, height, weight, systolic and diastolic blood pressure were measured in all participants. BMI was calculated as weight (kg)/ (height (m))². Serum levels of fasting glucose, triglycerides, total cholesterol, HDL-cholesterol, LDL-cholesterol, albumin, globulin, bilirubin, creatinine, urea, urate, HbA1c and C-reactive protein (CRP), as well as activities of aspartate aminotransferase, alanine aminotransferase, and y-glutamyltransferase were determined by using the VITROS auto analyzer 350 Chemistry System (Ortho-Clinical Diagnostics, Rochester, New York, USA). Serum insulin levels were measured by the Abbott AxSYM (Abbott Diagnostics, North Chicago, Illinois, USA) analyzer. HOMA IR index was calculated by using following formula: fasting insulin (mU/L) x fasting glucose (mmol/L)/22.5 (23).

Genotyping analysis

The Miller extraction protocol was used for the DNA extraction (24). Genotyping analysis for

rs11693809 (IVS1 +3341C > T, denoted intron 1 SNP) and rs2716610 (IVS17-228C > T, denoted intron 17 SNP) polymorphisms was performed with real-time PCR allelic discrimination on ABI PRISM with C_2096848_10 and C_16280532_10 assays, respectively (Applied Biosystems, Foster City, CA, USA). We double-genotyped twenty percent of all samples with 100% concordant results.

Statistical analysis

Chi-square (χ^2) and Fisher's exact tests (in the case where frequencies were less or equal to 5) were applied to examine differences in allele frequencies and genotype distributions between healthy controls and patients with MS. Significance of difference of biochemical and anthropometrical measurements according to genotypes of analyzed polymorphisms, sex and age were estimated by linear regression Corrections for

Table 1. Characteristics of the study participants

Parameter*	MS patients (n=43)	Controls (n=43)	p†
Age (years)	49 (40-56)	45 (41-51)	0.206
BMI (kg/m2)	33.0 (29.2-35.5)	24.7 (22.2-27.4)	< 0.001
Waist circumfe-	110 (96-120)	83 (78-90)	< 0.001
rence (cm)	110 (00 120)	05 (70 90)	-0.001
Systolic BP	143 (130-158)	120 (110-125)	< 0.001
(mm Hg) Diastolic BP			
(mm Hg)	90 (80-100)	78 (70-80)	< 0.001
Fasting insulin	10 ((0 0 12 0)	5 2 (6 4 10 1)	0.010
(mU/L)	10.6 (8.0-13.9)	7.3 (6.4-10.1)	0.010
Fasting glucose	8.4 (5.5-11.7)	5.0 (4.7-5.2)	< 0.001
(mmol/L)			
HOMA-IR	4.1 (2.7-6.1)	1.6 (1.4-2.3)	< 0.001
Blood HbA1c (%)	6.1 (5.5-7.2)	5.6 (4.8-6.0)	< 0.001
Total cholesterol	5.6 (5.1-6.3)	5.8 (5.2-6.5)	0.385
(mmol/L) LDL-cholesterol			
(mmol/L)	3.20 (2.60-4.01)	3.37 (2.87-4.19)	0.133
HDL-cholesterol			
(mmol/L)	1.07 (0.88-1.30)	1.67 (1.38-1.87)	< 0.001
Triglycerides	2.26 (1.77-3.27)	1.17 (0.76-1.45)	< 0.001
(mmol/L)	. ,		
CRP (mg/L)	5.0 (3.0-6.0)	1.3 (0.8-4.0)	< 0.001
Creatinine	82.5 (72.5-94.5)	96.0 (59.0-78.0)	< 0.001
(mmol/L)	5 05 (4 15 (12)	4 20 (2 70 5 40)	0.026
Urea (mmol/L)	5.05 (4.15-6.12)	4.30 (3.70-5.40)	0.036
Urate (mmol/L)	297.0 (239.0-333.0)	(
Albumin (g/L)	49.4 (43.0-52.7)	44.0 (42.0-48.1)	0.012
Globulin (g/L)	24.6 (21.4-30.7)	32.0 (28.0-33.0)	< 0.001
Bilirubin	12.9 (10.7-14.7)	11.5 (9.7-14.6)	0.302
(mmol/L) AST (IU/L)	26.0 (18.2-31.0)	26.0 (21.0-31.0)	0.593
AST (IU/L) ALT (IU/L)	27.0 (22.2-42.2)	23.0 (16.0-30.0)	0.595
. ,		. ,	0.017
GGT (IU/L)	23.5 (17.7-38.0)	17.5 (14.0-26.5)	0.298

*Values represent medians (lower-upper quartile); †significance of difference in Mann-Whitney test.

BMI, body mass index; BP, blood pressure; HOMA-IR, homeostasis model assessment insulin resistance index; LDL, low-density lipoprotein; HDL, high-density lipoprotein; hsCRP, high-sensitivity C-reactive protein; AST, aspartate aminotransferase; ALT, alanine aminotransferase; GGT – γ , glutamyl transferase γ multiple testing were performed by using Bonferroni correction for a total of two SNPs. A p value ≤ 0.05 was considered statistically significant. Linkage disequilibrium was calculated by using the 2LD program (25). Haplotype reconstruction was done with the PHASE program (26).

RESULTS

All analyzed parameters were significantly different between patients and control groups except total cholesterol, LDL cholesterol, and bilirubin levels, as well as aspartate aminotransferase (AST) and γ -glutamyltransferase activity (GGT) (Table 1).

Table 2. Allele and genotype frequencies for *LPIN1* gene polymorphisms*

Polymorp-			T-allele		T-allele	
hism		MS	frequ-	Controls	frequ-	p†
mam			ency		ency	
Intron	CC	16 (40.0%)		14 (33.3%)		
1SNP	CT	19 (47.5%)	0.36	23 (54.8%)	0,39	0.792
(rs11693809)	TT	5 (12.5%)		5 (11.9%)		
	Total	40		42		
	р	0.985		0.631		
Intron	CC	26 (65.0%)		32 (76.2%)		
17 SNP	CT	12 (30.0%)	0.20	9 (21.4%)	0.13	0.513
(rs2716610)	TT	2 (5.0%)		1 (2.4%)		
	Total	40		42		
	p‡	0.925		0.931		

*rs11693809: C>T and rs2716610: C>T; \dagger Significance of $\chi 2$ / Fisher's exact test for comparison of genotype frequencies between healthy controls and MS patients; $\ddagger p$ value for Hardy-Weinberg equilibrium.

Table 3. Effects of LPIN1 SNPs on biochemical and anthropometrical parameters in controls

$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		rs11693809 C>T					rs2716610 C>T				
	Parameter*	C/C	C/T + T/T	D (059/ CD +		"D	C/C	C/T + T/T	D (059/ CD+		
		(n=14)	(n=24)	B (95% CI) †	PT	рв	(n=29)	(n=10)	B (95% CI)†	Pf	рв
	BMI	23.3	25.1	2.156	0.1.41	0.000	25.4	21.0	-4.272	0.004	0.000
(cm) (72-85) (78-93) (-1.405, 11.740) 0.119 0.238 (80-93) (72-81) (-16-032, -3.243) 0.000 0.0008 Systolic BP 120 13.803 80.3 52.21 1.000 120 -5.752 0.364 0.728 Diastolic BP 75 80 2.255 0.541 1.000 (70-80) (67-82) (5.81/1, 2.722) 0.438 0.875 -0.974 0.805 1.000 Fasting insulin 7.3 6.9 -3.646 0.12 0.668 0.66 0.782 (4.874, 7.058, 7.014) 0.805 1.000 (mull.) (6.5-16 (4.77, 1 0.509, 0.213 0.55 1.000 (4.8-52) (4.3-53) (-1.66, 0.063 0.313 0.626 Blood IbA.1R 1.6 1.5 -0.880 0.89 0.178 1.42.31 (1.2-722) (-1.670, 0.564) 0.313 0.626 Blood IbA.1R 1.6 0.5 0.011 0.428 0.771 0.781 0.000 0.781 0.000	(kg/m2)	(20.5-25.4)	(22.2-28.7)	(-0.758, 5.071)	0.141	0.282	(22.9-28/.7)	(20.2-24.0)	(-7.104, -1.441)	0.004	0.008
(cm) (72-85) (78-93) (71-405, 11/40) (80-93) (72-81) (16-1032, 2343) (mm Hg) (101-137) (120-125) (8.272, 15.878) 0.21 1000 120 -5.752 0.974 (mm Hg) (66-87) (75-80) (5.21, 9.728) 0.511 1000 69 7.3 -1.095 0.438 0.876 (mm/L) (62-14.7) (5.8-7.7) (7.580, 0.201) 0.533 1.000 6.99 7.3 -1.095 0.438 0.876 (mm/L) (62-14.7) (5.8-7.7) (7.580, 0.201) 0.533 1.000 (48-5.2) (4.6 -0.128 0.057 0.114 HOMA-IR 1.6 1.5 -0.880 0.87 1.14-2.3 (12-2.2) (1.606, 0.010) 0.566 0.161 0.56 5.0 4.4 5.5 5.0 4.4 5.5 5.0 6.066 0.114 0.50 1.14 1.5 1.5 -0.566 0.609 1.000 1.50 1.5 5.0 5.0 0.609	Waist circumference	81	88	5.168	0.110	0 228	86	78	-9.637	0.004	0.008
The plot (101-137) (120-125) (-8.272, 15.878) 0.524 1.000 (102-120) (-18.509, 7.004) 0.364 0.728 Diastolic BP 75 80 2.255 0.541 1.000 (102-120) (-18.509, 7.004) 0.364 0.728 Fasting insulin 7.3 6.9 -3.646 0.068 0.136 (6.1-9) (6.0-9, 6) (-6.112, 2.72) 0.438 0.875 Fasting glucose 5.1 4.9 -0.089 0.553 1.000 (4.8-52) (4.3-5.3) (-0.066, 0.010) 0.114 HOMA-IR 1.6 1.5 -0.089 0.178 1.5 1.5 -0.566 0.066 0.133 0.626 Bibod HbAtc 5.6 5.6 0.161 0.78 1.000 (4.4-23) (1.2-22) (1.497, 0.58, 0.981) 0.308 0.878 1.000 Cital cholesterol 6.27.7 -0.334 0.357 0.114 5.9 6.0 0.199 0.609 1.000 IDL-cholesterol 1.72 1.6 0	(cm)	(72-85)	(78-93)	(-1.405, 11.740)	0.119	0.238	(80-93)	(72-81)	(-16-032, -3.243)	0.004	0.008
(mm Hg) (101-137) (120-125) (+2372, 15.878) (+20-130) (120-122) (+18, 50, 7, 004) (+18, 12, 12, 12, 12, 12, 12, 12, 12, 12, 12	Systolic BP	120	120	3.803	0 524	1 000	120	120		0 364	0 728
	(mm Hg)	()	()	(-8.272, 15.878)	0.524	1.000	()	()		0.504	0.720
					0 541	1 000				0.805	1 000
	(mm Hg)	()	()	()	0.5 11	1.000	()	()		0.005	1.000
	8				0.068	0.136				0 4 3 8	0.876
Manual (Mamol/L) (4.5-5.6) (4.7-5.1) (-0.392, 0.213) 0.553 1.000 (4.8-5.2) (4.3-5.3) (-0.666, 0.010) 0.057 0.114 HOMA-IR 1.6 1.5 -0.880 0.089 0.178 1.5 1.5 -0.566 0.0161 0.472 0.944 1.4.2.3) (12-22) (1.607, 0.564) 0.313 0.626 Biood HbA1c 5.6 5.6 0.161 0.472 0.944 (4.8-6.0) (5.0-5.8) (-0.420, 0.549) 0.788 1.000 C(%) (5.0-6.0) (5.1-6.4) (-1059, 0.392) 0.357 0.714 5.9 6.0 0.199 0.609 1.000 (mmol/L) (2.974, 85) (2.354, 18) (-0.904, 0.596) 1.000 (2.88, 446) (2.88, 446) (2.88, 446) (2.88, 446) (2.88, 446) (2.88, 446) (2.88, 446) (2.98, 454) (-0.679, 0.926) 1.000 Itriglycerides 1.15 1.12 -0.073 0.690 1.000 1.13 1.29 -0.008 0.968 1.000	()	()		()	0.000	0.150	()	()		0.100	0.070
(mmolL) (4.5-5.6) (4.7-5.1) (-0.392, 0.213) (4.8-5.2) (4.3-5.3) (-0.606, 0.010) HOMA-IR (1.3-1.1) (1.3-1.7) (-1.905, 0.144) 0.089 0.178 (1.5.1) (-0.566) 0.313 0.626 Blood HbA1c 5.6 5.6 0.161 0.472 0.944 5.6 5.6 0.005 0.788 1.000 Total cholesterol 6.2 5.7 -0.334 0.357 0.714 (5.1-6.4) (5.5-6.8) (-0.584) 0.609 1.000 (mmolL) (2.97-4.85) (2.93-4.18) (-0.904, 0.596) 0.771 0.164 (5.5-6.8) (-0.584, 0.981) 0.757 1.000 (mmolL) (1.48-2.03) (1.37-1.86) (-0.474, 0.064) 0.130 0.260 1.13 1.29 -0.008 0.914 (mmolL) (0.92-1.67) (0.67-1.44) (-0.439, 0.294) 0.469 1.001 1.1 1.01 0.241 0.448 0.277 0.554 (mmolL) (0.5-1.4) (0.94.7) (-0.582, 2.889 </th <th>00</th> <th></th> <th></th> <th></th> <th>0 553</th> <th>1 000</th> <th></th> <th></th> <th></th> <th>0.057</th> <th>0 114</th>	00				0 553	1 000				0.057	0 114
HOMA-IR (1.3-3.1) (1.3-1.7) (-1.905, 0.144) 0.089 0.178 (1.4-2.3) (1.2-2.2) (-1.697, 0.564) 0.313 0.626 Blood HbA1c 5.6 5.6 0.161 0.472 0.944 5.6 5.6 0.005 0.788 1.000 Total cholesterol 6.2 5.7 -0.334 0.357 0.714 (5.5-6.8) (-0.280, 0.910) 0.609 1.000 (mmol/L) (2.97-4.85) (2.93.4.18) (-0.904, 0.596) 0.771 1.000 2.884.46 (2.98-4.54) (-0.679, 0.926) 0.757 1.000 (mmol/L) (1.48-203) (1.37.1.86) (-0.474, 0.064) 0.130 0.260 1.13 1.29 -0.008 0.922 1.000 (mmol/L) (0.92-1.67) (0.67-1.44) (-0.439, 0.294) 0.16 0.333 0.48 0.17 1.154 0.370 1.13 1.29 -0.008 0.277 0.554 (mmol/L) (0.5-1.4) (0.94.77) (-0.582, 2.889 0.370 1.51 1.1 <	(mmol/L)	(· · · ·	()	0.000	1.000	· · · · ·			0.027	0.111
	HOMA-IR				0.089	0 1 7 8				0 313	0.626
		(0.009	0.170	· · · · ·			0.010	0.020
					0.472	0.944				0.788	1.000
(mmol/L) (5.3-7.0) (5.1-6.4) (1-059, 0.392) 0.357 0.714 (5.1-6.4) (5.5-6.8) (-0.584, 0.981) 0.609 1.000 LDL-cholesterol 3.81 3.35 -0.154 0.679 1.000 3.34 3.69 0.123 0.757 1.000 (mmol/L) (2.97-4.85) (2.93-4.18) (0.044, 0.064) 0.679 1.000 (2.88-4.46) (2.98-4.54) (-0.679, 0.926) 0.757 1.000 (mmol/L) (1.48-2.03) (1.37-1.86) (-0.474, 0.064) 0.130 0.260 1.13 1.29 -0.008 0.988 1.000 (mmol/L) (0.92-1.67) (0.67-1.44) (-0.439, 0.294) 0.690 1.000 (1.77-1.47) (0.61-1.49) (-0.401, 0.385) 0.277 0.554 (mmol/L) (0.5-1.4) (0.94-7) (-0.582, 2.889 0.870 0.15 1.1 -1.018 0.277 0.554 (mmol/L) (5.6-5.5) (3.85-4) (-0.70, -1.082) 0.278 0.873 0.000 2.85 2.440 0.299 0.058 0.029 0.058 (mmol/L) (2.5-5.5) <		(()			(()			
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$					0.357	0.714				0.609	1.000
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	()			())			(
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$					0.679	1.000				0.757	1.000
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	()			()			((
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $					0.130	0.260				0.922	1.000
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	()	(· /	()			· /				
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	0.				0.690	1.000				0.968	1.000
	()		· /	()			· · · · ·	· /			
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $					0.185	0.370				0.277	0.554
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		((()		
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $					0.242	0.484				0.029	0.058
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		(· /	()			()				
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$					0.671	1.000				0.081	0.162
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	()		(· · · ·			(()	()		
Albumin 44.5 44.0 -1.690 0.208 0.416 44.0 44.5 0.619 0.669 1.000 (g/L) $(42.7-49.1)$ $(42.2-47.4)$ $(-4.365-0.985)$ 0.208 0.416 $(42.0-47.8)$ $(42.7-47.4)$ $(-2.302, 3.540)$ 0.669 1.000 Globulin 30.0 32.0 2.900 0.110 0.220 32.0 31.5 -0.075 0.970 1.000 (g/L) $(25.4-33.0)$ $(28.7-34.5)$ $(-0.689-6.488)$ 0.110 0.220 32.0 31.5 -0.075 0.970 1.000 Bilirubin 9.6 12.5 2.488 0.088 0.176 $(9.6-15.6)$ $(7.7-11.9)$ $(4.578, 1.777)$ 0.377 0.754 AST 22.0 26.0 1.358 0.491 0.982 26.0 23.5 -1.147 0.587 1.000 ALT 21.0 24.0 0.343 0.928 1.000 $(14.0-28.5)$ $(16.7-35.2)$ $(-3.894, 12.208)$ 0.301 0.602 GGT 16.0 18.0 -5.483 0.514 1.000 17.0 18.5 15.814 0.072 0.144					0.873	1.000				0.902	1.000
	()	()	(()	()	()		
Globulin 30.0 32.0 2.900 0.110 0.220 32.0 31.5 -0.075 0.970 1.000 (g/L) (25.4-33.0) (28.7-34.5) (-0.689-6.488) 0.110 0.220 (28.8-34.0) (27.4-34.0) (-4.062, 3.913) 0.970 1.000 Bilirubin 9.6 12.5 2.488 0.088 0.176 (9.6-15.6) (7.7-11.9) (-4.578, 1.777) 0.377 0.754 AST 22.0 26.0 1.358 0.491 0.982 26.0 23.5 -1.147 0.587 1.000 (IU/L) (20.5-30.0) (21.5-30.7) (-2.611-5.326) 0.928 1.000 (23.0 22.0 4.157 0.587 1.000 ALT 21.0 24.0 0.343 0.928 1.000 (14.0-28.5) (16.7-35.2) (-3.894, 12.208) 0.301 0.602 GGT 16.0 18.0 -5.483 0.514 1.000 17.0 18.5 15.814 0.072 0.144 (IU/L)					0.208	0.416				0.669	1.000
		(· /			()	()	()		
Bilirubin (mmol/L) 9.6 12.5 2.488 0.088 0.176 11.7 10.6 -1.401 0.377 0.754 AST 22.0 26.0 1.358 0.088 0.176 (9.6-15.6) (7.7-11.9) (-4.578, 1.777) 0.377 0.754 AST 22.0 26.0 1.358 0.491 0.982 26.0 23.5 -1.147 0.587 1.000 ALT 21.0 24.0 0.343 0.928 1.000 21.5-30.5) (15.7-35.2) (-3.894, 12.208) 0.301 0.602 GGT 16.0 18.0 -5.483 0.514 1.000 17.0 18.5 15.814 0.072 0.144 (U/L) (13.7-28.7) (15.0-26.0) (-22.380-11.414) 0.514 1.000 17.0 18.5 15.814 0.072 0.144					0.110	0.220				0.970	1.000
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		(((((
AST 22.0 26.0 1.358 0.491 0.982 26.0 23.5 -1.147 0.587 1.000 (IU/L) $(20.5-30.0)$ $(21.5-30.7)$ $(-2.611-5.326)$ 0.491 0.982 $(21.5-30.5)$ $(19.5-30.0)$ $(-5.399, 3.105)$ 0.587 1.000 ALT 21.0 24.0 0.343 0.928 1.000 23.0 22.0 4.157 0.301 0.602 (IU/L) $(11.7-29.0)$ $(16.0-29.7)$ $(-7.313-8.000)$ 0.928 1.000 17.0 18.5 15.814 0.072 0.144 (IU/L) $(13.7-28.7)$ $(15.0-26.0)$ $(-22.380-11.414)$ 0.514 1.000 $(15.0-25.5)$ $(13.7-35.5)$ $(-1.500, 33.127)$					0.088	0.176				0.377	0.754
		(· · · ·	· /			()		())		
ALT 21.0 24.0 0.343 0.928 1.000 23.0 22.0 4.157 0.301 0.602 (IU/L) $(11.7-29.0)$ $(16.0-29.7)$ $(-7.313-8.000)$ 0.928 1.000 $(14.0-28.5)$ $(16.7-35.2)$ $(-3.894, 12.208)$ GGT 16.0 18.0 -5.483 0.514 1.000 17.0 18.5 15.814 0.072 0.144 (IU/L) $(13.7-28.7)$ $(15.0-26.0)$ $(-22.380-11.414)$ 0.514 1.000 $(15.0-25.5)$ $(13.7-35.5)$ $(-1.500, 33.127)$					0.491	0.982				0.587	1.000
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		((
GGT 16.0 18.0 -5.483 0.514 1.000 17.0 18.5 15.814 0.072 0.144 (IU/L) (13.7-28.7) (15.0-26.0) (-22.380-11.414) 0.514 1.000 17.0 18.5 15.814 0.072 0.144					0.928	1.000				0.301	0.602
$(IU/L) \qquad (13.7-28.7) (15.0-26.0) (-22.380-11.414) \xrightarrow{0.514 \ 1.000} (15.0-25.5) (13.7-35.5) (-1.500, 33.127) \xrightarrow{0.072 \ 0.144}$											
					0.514	1.000				0.072	0.144
*Values concept modiane (lower upper guartile): *Effects (unstandardized exafficients B) and a values were assessed using multiple linear regree	<u> </u>						/	/			

*Values represent medians (lower-upper quartile); †Effects (unstandardized coefficients B) and p values were assessed using multiple linear regression adjusted for age and gender, under dominant genetic model; pB, adjusted by using Bonferroni correction for two SNPs;

BMI, body mass index; BP; blood pressure; HOMA-IR, homeostasis model assessment insulin resistance index; LDL, low-density lipoprotein; HDL, high-density lipoprotein; hsCRP, high-sensitivity C-reactive protein; AST, aspartate aminotransferase; ALT, alanine aminotransferase; GGT-7; glutamyltransferase;

		rs11	693809 C>T				rs2	716610 C>T					
Parameter*	C/C	C/T + T/T	D (ARA) CD I		pB	C/C	C/T + T/T	D (ARA) CD I					
	(n=16)	(n=24)	B (95% CI)†	p†	•	(n=26)	(n=14)	B (95% CI)†	p†	pВ			
BMI	33.5	33.0	0.280			33.5	33.0	-1.825					
(kg/m2)	(29.3-36.4)	(26.9-35.3)	(-3.845, 4.404)	0.891	1.000	(28.9-36.9)	(25.7-35.1)	(-5.904, 2.253)	0.370	0.74			
Waist circumference	111	111	-5.482			112	110	-6.339					
(cm)	(103 - 117)	(96-123)	(-17.623, 6.659)	0.365	0.730	(106-120)	(87-122)	(-18.423, 5.746)	0.293	0.58			
Systolic BP	145	140	-6.002			145	140	-7.799					
(mm Hg)	(140-160)	(130-155)	(-20.321, 8.317)	0.398	0.796	(140-160)	(130-150)	(-21.990, 6.392)	0.270	0.54			
Diastolic BP	95	90	-4.814			92	85	-6.985					
(mm Hg)	(81-100)	(80-97)	(-14.276, 4.647)	0.307	0.614	(81-100)	(70-97)	(.16.248, 2.277)	0.134	0.26			
Fasting insulin	10.3	11.5	0.381			10.4	11.3	-0.869					
(mU/L)	(7.3-12.6)	(8.9-16.4)	(-4.951, 5.712)	0.885	1.000	(7.4-12.9)	(9.0-16.0)	(-6.191, 4.454)	0.741	1.00			
Fasting glucose	11.0	6.2	-2.315			9.0	6.1	-2.118					
(mmol/L)	(7.2-14.0)	(5.4-9.8)	(-5.305, 0.675)	0.125	0.250	(5.8-11.9)	(5.5-10.3)	(-5.124, 0.888)	0.162	0.32			
(millor/L)	(7.2-14.0) 4.3	(3.4-9.8)	0.088			4.3	(3.3-10.3)	(-3.124, 0.888) -1.047					
HOMA-IR	(3.6-5.6)	(2.6-7.7)	(-1.909, 2.084)	0.929	1.000	(3.3-8.0)	(2.3-5.6)	(-3.006, 0.912)	0.284	0.50			
Blood HbA1c	(3.6-3.6) 6.9	(2.0-7.7)	-1.082			(3.3-8.0) 6.4	(2.3-3.6)	-0.223					
				0.024	0.048				0.653	1.00			
(%) Total cholesterol	(5.9-7.7) 5.6	(5.2-6.6) 5.6	(-2.010, -0.154) 0.142			(5.5-7.4) 5.7	(5.5-6.5) 5.5	(-1.222, 0.775) -0.228					
				0.711	1.000				0.551	1.00			
(mmol/L)	(5.1-6.0)	(5.0-6.4)	(-0.629, 0.913)			(5.1-6.2)	(4.5-6.3)	(-0.996, 0.549)					
LDL-cholesterol	3.02	3.31	0.309	0.451	0.902	3.16	3.05	-0.016	0.969	1.00			
(mmol/L)	(2.56-3.28)		(-0.515, 1.134)				(2.01-4.18)	(-0.848, 0.815)					
HDL-cholesterol	1.11	1.00	0.043	0.701	1.000	1.20	0.91	-0.228	0.033	0.0			
(mmol/L)	(0.83-1.42)	()	(-0.181, 0.266)			()	(0.83-1.13)	(-0.438, -0.019)					
Triglycerides	2.38	2.24	-0.595	0.177	0.354	2.22	2.37	-0.112	0.801	1.00			
(mmol/L)		(1.88-3.69)	(-1.473, 0.283)				(1.69-4.03)	(-1.013, 0.788)					
hsCRP	5.0	5.0	-0.103	0.910	1.000	5.0	5.0	0.233	0.798	1.00			
(mg/L)	(3.2-6.0)	(2.2-6.7)	(-1.946, 1.740)			(3.0-6.0)	(2.0-7.2)	(-1.608, 2.074)					
Creatinine	83.0	83.0	2.708	0.723	1.000	82.0	89.0	-0.377	0.961	1.00			
(mmol/L)	(()	(-12.677, 18.093)	0.720	1.000			(-15.788, 15.034)	0.901	1.00			
Urea	4.9	5.3	0.444	0.437	0.874	5.0	5.0	-0.110	0.848	1.00			
(mmol/L)	(3.9-5.8)	(4.66.2)	(-0.703, 1.590)	0.457	0.074	(4.3-6.0)	(3.9-6.5)	(-1.266, 1.047)	0.040	1.00			
Urate	275	325	38.578	0.185	0.370	295	316	-0.178	0.995	1.00			
(mmol/L)	(232-336)	(294-338)	(-19.443, 96.600)	0.165	0.570	(246-337)	(278-337)	(-59.775, 59.419)	0.995	1.00			
Albumin	49.9	48.0	-0.799	0.686	1.000	50.5	46.4	-2.595	0.184	0.30			
(g/L)	(45.1-51.8)	(43.0-53.3)	(-4.794, 3.195)	0.080	1.000	(43.7-52.9)	(41.7-51.3)	(-6.488, 1.299)	0.184	0.50			
Globulin	26.0	23.5	-1.053	0 576	1.000	24.6	24.6	-0.464	0.806	1.0			
(g/L)	(21.6-30.3)	(21.4-32.0)	(-4.852, 2.746)	0.576	1.000	(21.5-30.2)	(19.7-33.2)	(-4.277, 3.350)	0.800	1.00			
Bilirubin	13.3	12.0	-2.408	0.151	0.302	13.3	12.0	-2.690	0.107	0.2			
(mmol/L)	(12.4-15.6)	(9.1-14.0)	(-5.738, 0.921)	0.131	0.302	(10.9-15-9)	(10.0-13.6)	(-5.993, 0.613)	0.107	0.2			
AST	22.0	26.0	3.968	0 272	0.744	26.0	22.5	-6.080	0.169	0.2			
(IU/L)	(17.0-28.0)	(19.2-31.7)	(-4.941, 12.876)	0.372	0.744	(19.5-31.5)	(18.0-30.2)	(-14.845, 2.686)	0.168	0.3.			
ALT	25.0	27.0	-0.843	0.004	1 000	33.0	23.5	-15.079	0.007	0.0			
(IU/L)	(22.0-35.0)	(22.2-48.0)	(-12.511, 10.826)	0.884	1.000	(23.0-50.5)	(20.0-33.0)	(-25.539, -4.619)	0.006	0.0			
GGT	15.0	25.0	-1.335	0.07/	1 000	23.0	22.5	-11.987	0.154	0.2			
(IU/L)	(8 75-27 5)	(16.0-38.0)	(-18.571, 15.900)	0.876	1.000	(12.0-38.0)	(9.5 - 31.2)	(-28.694, 4.721)	0.154	0.30			

Table 4. Effects of LPIN1 SNPs on	biochemical and	anthropometrical	parameters in the patients

*Values represent medians (lower-upper quartile);

†Effects (unstandardized coefficients B) and p values were assessed using multiple linear regression adjusted for age and gender, under dominant genetic model; pB adjusted by using Bonferroni correction for two SNPs;

BMI, body mass index; BP; blood pressure; HOMA-IR, homeostasis model assessment insulin resistance index; LDL, low-density lipoprotein; HDL, high-density lipoprotein; hsCRP, high-sensitivity C-reactive protein; AST, aspartate aminotransferase; ALT, alanine aminotransferase; GGT-γ; glutamyltransferase;

Table 5. Haplotype frequencies and diplotypes of LPIN1 gene in MS patients and healthy controls

Haplotype (intron 1 SNP-intron 17 SNP)*	Frequency		Distance	Diplotype No	No. of subjects	
	MS patients	Con- trols	– Diplotype		MS pa- tients	Con- trols
CC	0.548	0.505	CC/CC	1	14	8
СТ	0.102	0.116	CC/CT	2	3	4
TC	0.252	0.344	CC/TC	3	9	12
TT	0.098	0.035	CC/TT	4	8	-
			CT/CT	5	-	1
			CT/TC	6	-	3
			CT/TT	7	2	-
			TC/TC	8	3	4
			TC/TT	9	2	1

*corresponding to the rs11693809: C>T (intron SNP1) and rs2716610: C>T (intron SNP17) polymorphism

Allele frequencies were in Hardy-Weinberg equilibrium for both, patients and control subjects (p>0.05). However, no significant differences in analyzed genotype frequencies were found between patients and healthy controls (Table 2).

In a control group, after Bonferroni correction, rs11693809: C>T polymorphism did not show significant association with any biochemical or anthropometrical measurements. However, the carriers of T allele (CT + TT) of another analyzed polymorphism of *LPIN1* gene, rs2716610: C>T, had significantly lower BMI (p=0,008), waist circumference (p=0.008), and tendency of association with lower creatinine levels (p=0.058), as compared to the carriers of a wild type allele (CC) (Table 3).

In group of MS patients, the carriers of T allele (CT + TT) of rs11693809: C>T polymorphism had significantly lower blood HbA_{1c} (%) (p=0.048), as compared to the carriers of a wild type allele (CC). The carriers of T allele (CT + TT) of rs2716610: C>T polymorphism had significant lower ALT activity (p=0.012) and tendency of association with lower HDL cholesterol levels (p=0.066), as compared to the carriers of a wild type allele (CC). As shown in Table 4, no association was found for both analyzed *LPIN1* gene polymorphisms with anthropometrical measurements (body mass index (BMI) and waist circumference).

The selected *LPIN1* variants, rs11693809: C>T and rs2716610: C>T were in a weak linkage disequilibrium (D'=0.261). No significant differences in distribution of haplotype frequencies between patients with MS and control subjects were demonstrated. Furthermore, an association of *LPIN1* haplotypes with biochemical and anthropometrical parameters was also tested. In control group, the carriers of CC haplotype had significantly higher plasma insulin (p=0.024), higher glucose levels (p=0.036) and higher HOMA IR index (p=0.024) as compared to the carriers of CT haplotype. No significant associations of *LPIN1* haplotypes with traits of MS were found in patients group (Table 5).

DISCUSSION

Members of the lipin protein family have a newly discovered enzymatic role in triglyceride and phospholipid biosynthesis as a phosphatidate phosphatase, and act also as inducible transcriptional coactivators in conjunction with peroxisome proliferator-activated receptor c (PPARc) coactivator-1a and PPARa (13). Through these activities, the founding member of the family, lipin-1, influences lipid metabolism and glucose homeostasis (13).

Results of our study showed significant association of rs2716610: C>T genetic variant with body mass index (BMI) and waist circumference. No significant associations between disease-associated traits and rs11693809: C>T were found.

Since the majority of patients participating in this study were using medications (antihypertensive,

glucose-lowering or lipid-lowering drugs), the influence of *LPIN1* gene polymorphisms on the most of selected biochemical parameters in MS patients should be interpreted cautiously. However, it was reasonable to analyze the effects of *LPIN1* gene variations on the BMI and waist circumference in these patients.

The selected genotype-phenotype analysis showed that, the mutant T allele of rs11693809: C>T polymorphism was associated with lower blood HbA₁ in a group of MS patients. T allele of another analyzed polymorphism in our study, rs2716610: C>T, was associated with lower BMI and waist circumference, and creatinine levels in controls, and lower HDL cholesterol levels and lower ALT activity in a group of MS patients.

Only a few studies analyzed association of LPIN1 gene polymorphisms (rs11693809: C>T and rs2716610: C>T) with metabolic syndrome (17,22,27). Suviolahti et al. analyzed seven polymorphisms in the LPIN1 gene. They found an association of rs11693809: C>T polymorphism with the insulin levels. In addition, rs11693809: C>T and rs2716610: C>T polymorphism were associated with BMI in lean males. Since an association of LPIN1 variants with BMI in the lean or obese females was not found, the association of LPIN1 alleles with BMI appeared to be sex specific (17). Mlinar et al., in their study tested an association of LPIN1 polymorphisms (rs11693809: C>T and rs2716610: C>T) with polycystic ovary syndrome (PCOS) (28). Their results showed that mutated T allele of rs11693809: C>T polymorphism was associated with lower plasma LDL-cholesterol levels in controls, with lower glucose levels after OGTT in the PCOS patients, and with lower insulin levels and HOMA-IR in nonobese PCOS patients. These results suggest a protective role of mutated T allele of rs11693809: C>T polymorphism against development of IR and dyslipidemia. Mutated allele T of another analyzed polymorphism in this study (28), rs2716610: C>T, showed an association with higher triglyceride levels in control subjects, suggesting a negative effect of this polymorphism on the metabolic phenotype. Wiedmann at al. in their study analyzed an association of 15 genetic variants of LPIN1 gene, including rs2716610: C>T, with metabolic phenotype. Their results showed the borderline significant

association of this polymorphism (rs2716610: C>T) with higher plasma triglyceride levels (22). In our recent study we have also tested an association of LPIN1 and PPARG variants with biochemical and anthropometrical parameters in patients with MS and type 2 diabetes. Interestingly, results of this study showed that mutated T allele of rs11693809: C>T polymorphism was associated with higher insulin levels in patients with MS and type 2 diabetes (27). Neither of these studies analyzed an association of two LPIN1 variants (rs11693809: C>T and rs2716610: C>T) with the biochemical parameters, including albumin, globulin, creatinine, urea and uric acid levels, as well as ALT, AST and GGT activity that we tested in the current study. The mechanism of an association of LPIN1 polymorphisms with above biochemical parameters is not completely understood. However, many previous studies demonstrated an association of selected biochemical parameters, which are known as possible markers, with increased risk of development Type 2 diabetes (T2D) (29-33). Recently published studies found a positive correlation between higher creatinine and urea levels with increased risk for T2D development (32-33). Furthermore, other studies found a correlation of higher activity of liver enzymes (AST, ALT or GGT) with increased risk for MS and T2D development, or their association with obesity and insulin resistance (29-31).

Effects of two polymorphisms of *LPIN1* gene (rs11693809: C>T and rs2716610: C>T) on metabolic phenotype was further tested by the haplotype analyses. An association of rs2716610: C>T polymorphism with the lower BMI and waist circumference, and protective role of this polymorphism were confirmed by this analysis. Another analyzed polymorphism, rs11693809: C>T, did not show any association disease-associated traits, that is also confirmed by haplotype analysis. In control subjects, carriers of CT haplotype had significant lower insulin levels, glucose levels and HOMA IR levels as compa-

REFERENCE

- 1. Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. Diabetes 1988; 37:1595-607.
- DeFronzo RA, Ferrannini E. Insulin resistance. A multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. Diabetes Care 1991; 14:173-94.

red to the carriers of CC haplotype. Thus, these findings suggested that non-mutated first locus, and mutated second locus (CT haplotype) decreased metabolic risk, while non-mutated first and second locus (CC haplotype) increased metabolic risk. These positive findings have to be replicated in a larger cohort of subjects.

In conclusion, the reason why we have chosen these two SNPs of LPIN1 gene is because previous studies have shown association with metabolic phenotype, but with opposite results, prompting us to examine impact of these polymorphisms of LPIN1 gene in development of metabolic syndrome in the population of Bosnia and Herzegovina. These two polymorphisms did not cover the role of all variation of LPIN1 gene, which is a limitation of our study.

Results of our study showed that rs2716610: C>T decrease the risk of MS, while rs11693809: C>T did not show any association with risk factors of MS. An association of rs2716610: C>T polymorphism with lower BMI and waist circumference suggest that this genetic variant of *LPIN1* gene could have a protective role against development of metabolic syndrome. Future investigations including larger cohorts of subjects, genes and larger number of SNPs, as well as studies on other populations, are needed to substantiate these findings. Illuminating the mechanism and pathogenesis of this complex disorder, might lead to effective treatment, and also to predict individual's risk of developing of metabolic syndrome.

ACKNOWLEDGEMENTS

We greatly appreciate the technical assistance of Mr. Nermin Kotoric, Mr. Hajrudin Mujic and Ms. Elma Topić.

FUNDING

No specific funding was received for this study.

TRANSPARENCY DECLARATION

Competing interests: None to declare

- Kaplan NM. The deadly quartet. Upper-body obesity, glucose intolerance, hypertriglyceridemia, and hypertension. Arch Intern Med 1989; 149:1514-20.
- Eckel RH, Grundy SM, Zimmet PZ. The metabolic syndrome. Lancet 2005; 365:1415-28.
- Grundy SM. Metabolic syndrome pandemic. Arterioscler Thromb Vasc Biol 2008; 28:629-36.

- Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, Fruchart JC, James WP, Loria CM, Smith SC, Jr. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 2009; 120:1640-5.
- Grundy SM, Cleeman JI, Daniels SR, Donato KA, Eckel RH, Franklin BA, Gordon DJ, Krauss RM, Savage PJ, Smith SC, Jr., Spertus JA, Costa F. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. Circulation 2005; 112:2735-52.
- Donkor J, Sariahmetoglu M, Dewald J, Brindley DN, Reue K. Three mammalian lipins act as phosphatidate phosphatases with distinct tissue expression patterns. J Biol Chem 2007; 282:3450-7.
- Rankinen T, Zuberi A, Chagnon YC, Weisnagel SJ, Argyropoulos G, Walts B, Perusse L, Bouchard C. The human obesity gene map: the 2005 update. Obesity (Silver Spring) 2006; 14:529-644.
- Peterfy M, Phan J, Xu P, Reue K. Lipodystrophy in the fld mouse results from mutation of a new gene encoding a nuclear protein, lipin. Nat Genet 2001; 27:121-4.
- Peterfy M, Phan J, Reue K. Alternatively spliced lipin isoforms exhibit distinct expression pattern, subcellular localization, and role in adipogenesis. J Biol Chem 2005; 280:32883-9.
- 12. Phan J, Reue K. Lipin, a lipodystrophy and obesity gene. Cell Metab 2005; 1:73-83.
- 13. Reue K, Dwyer JR. Lipin proteins and metabolic homeostasis. J Lipid Res 2009; 50 Suppl:S109-14.
- 14. Donkor J, Sparks LM, Xie H, Smith SR, Reue K. Adipose tissue lipin-1 expression is correlated with peroxisome proliferator-activated receptor alpha gene expression and insulin sensitivity in healthy young men. J Clin Endocrinol Metab 2008; 93:233-9.
- Croce MA, Eagon JC, LaRiviere LL, Korenblat KM, Klein S, Finck BN. Hepatic lipin 1beta expression is diminished in insulin-resistant obese subjects and is reactivated by marked weight loss. Diabetes 2007; 56:2395-99.
- van Harmelen V, Ryden M, Sjolin E, Hoffstedt J. A role of lipin in human obesity and insulin resistance: relation to adipocyte glucose transport and GLUT4 expression. J Lipid Res 2007; 48:201-6.
- Suviolahti E, Reue K, Cantor RM, Phan J, Gentile M, Naukkarinen J, Soro-Paavonen A, Oksanen L, Kaprio J, Rissanen A, Salomaa V, Kontula K, Taskinen MR, Pajukanta P, Peltonen L. Cross-species analyses implicate Lipin 1 involvement in human glucose metabolism. Hum Mol Genet 2006; 15:377-86.
- Loos RJ, Rankinen T, Perusse L, Tremblay A, Despres JP, Bouchard C. Association of lipin 1 gene polymorphisms with measures of energy and glucose metabolism. Obesity (Silver Spring) 2007; 15:2723-32.
- Cao H, Hegele RA. Identification of single-nucleotide polymorphisms in the human LPIN1 gene. J Hum Genet 2002; 47:370-2.

- 20. Fawcett KA, Grimsey N, Loos RJ, Wheeler E, Daly A, Soos M, Semple R, Syddall H, Cooper C, Siniossoglou S, O'Rahilly S, Wareham NJ, Barroso I. Evaluating the role of LPIN1 variation in insulin resistance, body weight, and human lipodystrophy in U.K. Populations. Diabetes 2008; 57:2527-33.
- Ong KL, Leung RY, Wong LY, Cherny SS, Sham PC, Lam TH, Lam KS, Cheung BM. Association of a polymorphism in the lipin 1 gene with systolic blood pressure in men. Am J Hypertens 2008; 21:539-45.
- 22. Wiedmann S, Fischer M, Koehler M, Neureuther K, Riegger G, Doering A, Schunkert H, Hengstenberg C, Baessler A. Genetic variants within the LPIN1 gene, encoding lipin, are influencing phenotypes of the metabolic syndrome in humans. Diabetes 2008; 57:209-17.
- 23. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia 1985; 28:412-9.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res 1988; 16:1215.
- Zhao JH. 2LD, GENECOUNTING and HAP: Computer programs for linkage disequilibrium analysis. Bioinformatics 2004; 20:1325-6.
- Stephens M, Smith NJ, Donnelly P. A new statistical method for haplotype reconstruction from population data. Am J Hum Genet 2001; 68:978-89.
- 27. Bego T, Dujic T, Mlinar B, Semiz S, Malenica M, Prnjavorac B, Ostanek B, Marc J, Causevic A. Association of PPARG and LPIN1 gene polymorphisms with metabolic syndrome and type 2 diabetes. Med Glas Ljek komore Zenicko-doboj kantona 2011; 8:76-83.
- Mlinar B, Ferk P, Pfeifer M, Gersak K, Marc J. Lipin 1 gene polymorphisms in polycystic ovary syndrome. Horm Metab Res 2011; 43:427-32.
- 29. Forlani G, Di Bonito P, Mannucci E, Capaldo B, Genovese S, Orrasch M, Scaldaferri L, Di Bartolo P, Melandri P, Dei Cas A, Zavaroni I, Marchesini G. Prevalence of elevated liver enzymes in Type 2 diabetes mellitus and its association with the metabolic syndrome. J Endocrinol Invest 2008; 31:146-52.
- 30. Marchesini G, Avagnina S, Barantani EG, Ciccarone AM, Corica F, Dall'Aglio E, Dalle Grave R, Morpurgo PS, Tomasi F, Vitacolonna E. Aminotransferase and gamma-glutamyltranspeptidase levels in obesity are associated with insulin resistance and the metabolic syndrome. J Endocrinol Invest 2005; 28:333-9.
- 31. Zhang Y, Lu X, Hong J, Chao M, Gu W, Wang W, Ning G. Positive correlations of liver enzymes with metabolic syndrome including insulin resistance in newly diagnosed type 2 diabetes mellitus. Endocrine 2010; 38:181-7.
- 32. Lal SS, Sukla Y, Singh A, Andriyas AE, Lall MA. Hyperuricemia, high serum urea and hypoproteinemia are the risk facror for diabetes. Asian Journal of Medical Sciences 2009; 1:33-4.
- 33. Idonije BO, Festus O, Oluba MO. Plasma glucose, creatinine and urea levels in type 2 diabetic patients attending a Nigerian Teaching Hospital. Research Journal of Medical Sciences 2011; 5:1-3.

Povezanost varijacija *LPIN1* gena s markerima metaboličkog sindroma u populaciji Bosne i Hercegovine

Tamer Bego¹, Tanja Dujić¹, Barbara Mlinar², Sabina Semiz^{1,4}, Maja Malenica¹, Besim Prnjavorac^{3,6}, Barbara Ostanek², Janja Marc², Anida Čaušević-Ramoševac⁵, Adlija Čaušević¹

¹Katedra za biohemiju i kliničke analize, Farmaceutski fakultet, Univerzitet u Sarajevu, Sarajevo, Bosna i Hercegovina; ²Katedra za kliničku biohemiju, Farmaceutski fakultet, Univerzitet u Ljubljani, Ljubljana, Slovenija; ³Opća bolnica u Tešnju, Tešanj, ⁴Fakultet za inženjering i prirodne nauke, Internacionalni univerzitet u Sarajevu, Sarajevo, ⁵Bosnalijek d. d, Farmaceutska i hemijska industrija, Sarajevo, ⁶Katedra za patofiziologiju, Farmaceutski fakultet, Univerzitet u Sarajevu, Sarajevo, Sarajevo, Bosna i Hercegovina

SAŽETAK

Cilj Istražiti povezanost dvije varijante *LPIN1* gena s glavnim karakteristikama metaboličkog sindroma (MS) (opseg struka, indeks tjelesne mase, krvni pritisak, trigliceridi, HDL holesterol i glukoza) u populaciji Bosne i Hercegovine.

Metode Studija je uključila 43 pacijenta s metaboličkim sindromom i 43 zdrava ispitanika (kontrole) iz Opće bolnice u Tešnju. Varijante *LPN1* gena (rs11693809: C>T i rs2716610: C>T) analizirane su PCR metodom (*real time* PCR).

Rezultati Kod kontrolnih ispitanika, polimorfizam *LPIN1* gena, rs2716610: C>T, sa statistički značajnom razlikom bio je povezan s nižim vrijednostima indeksa tjelesne mase (ITM) (p=0.008) i opsega struka (p=0.008). Drugi analizirani genski polimorfizam, rs11693809: C>T, pokazao je povezanost s nižim vrijednostima HbA1c (p=0.048) u skupini pacijenata s MS-om.

Zaključak Rezultati studije sugerišu da bi rs2716610: C>T polimorfizam *LPIN1* gena mogao imati zaštitnu ulogu u razvoju metaboličkog sindroma, dok bi polimorfizam rs11693809: C>T mogao imati ulogu u kontroli glukoze kod pacijenata s MS-om.

Ključne riječi: metabolički sindrom, LPIN1 gen, marker, varijacije gena